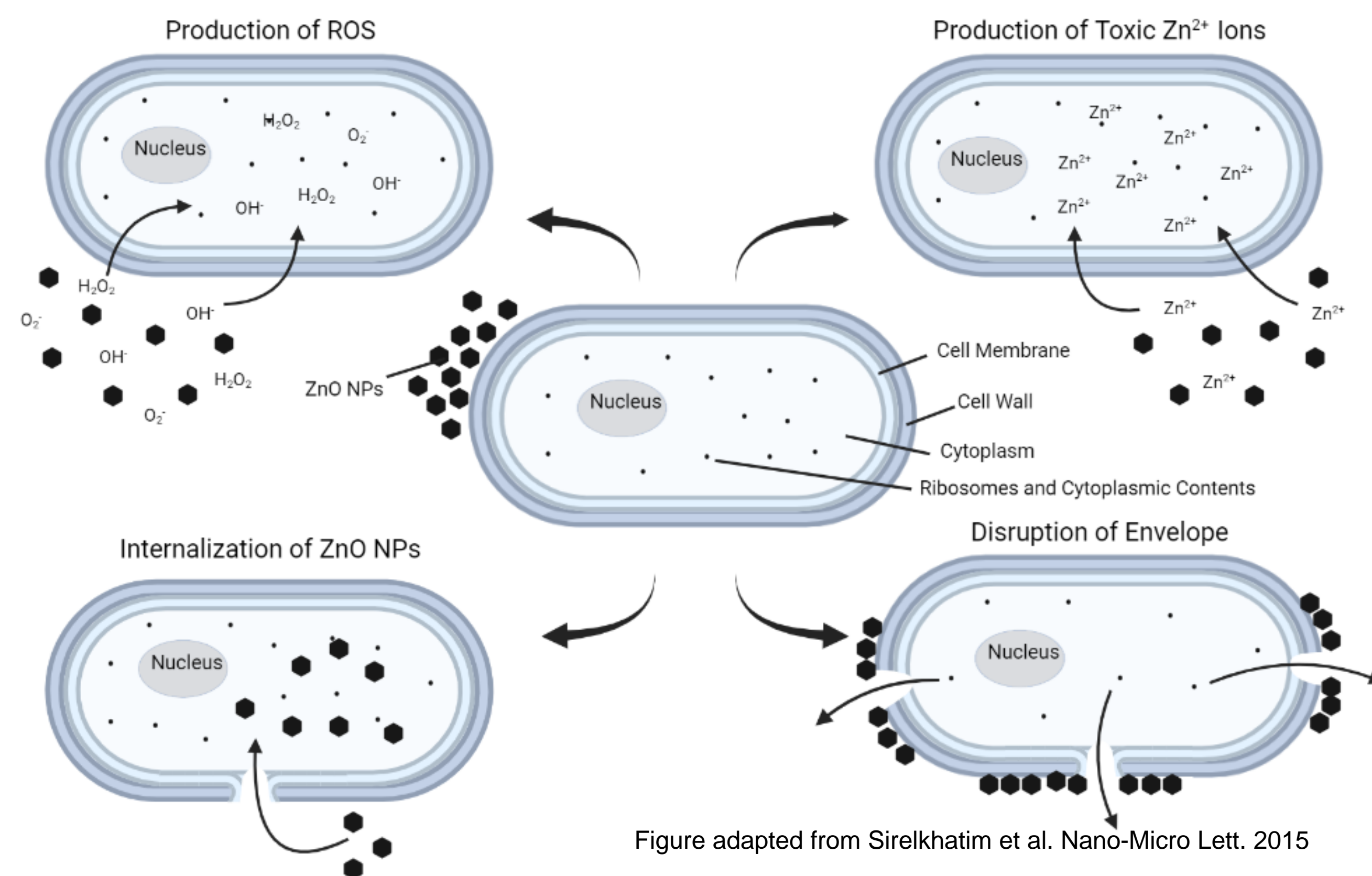


Characterization of Antibacterial Mechanisms of Zinc Oxide in *Staphylococcus aureus*

Alexander Caron¹, John Reeks², Dustin Johnson¹, Iman Ali¹, Michael Delgado¹, Shauna M. McGillivray¹
¹Texas Christian University, ²Institute of Low Temperature and Structure Research Polish Academy of Sciences

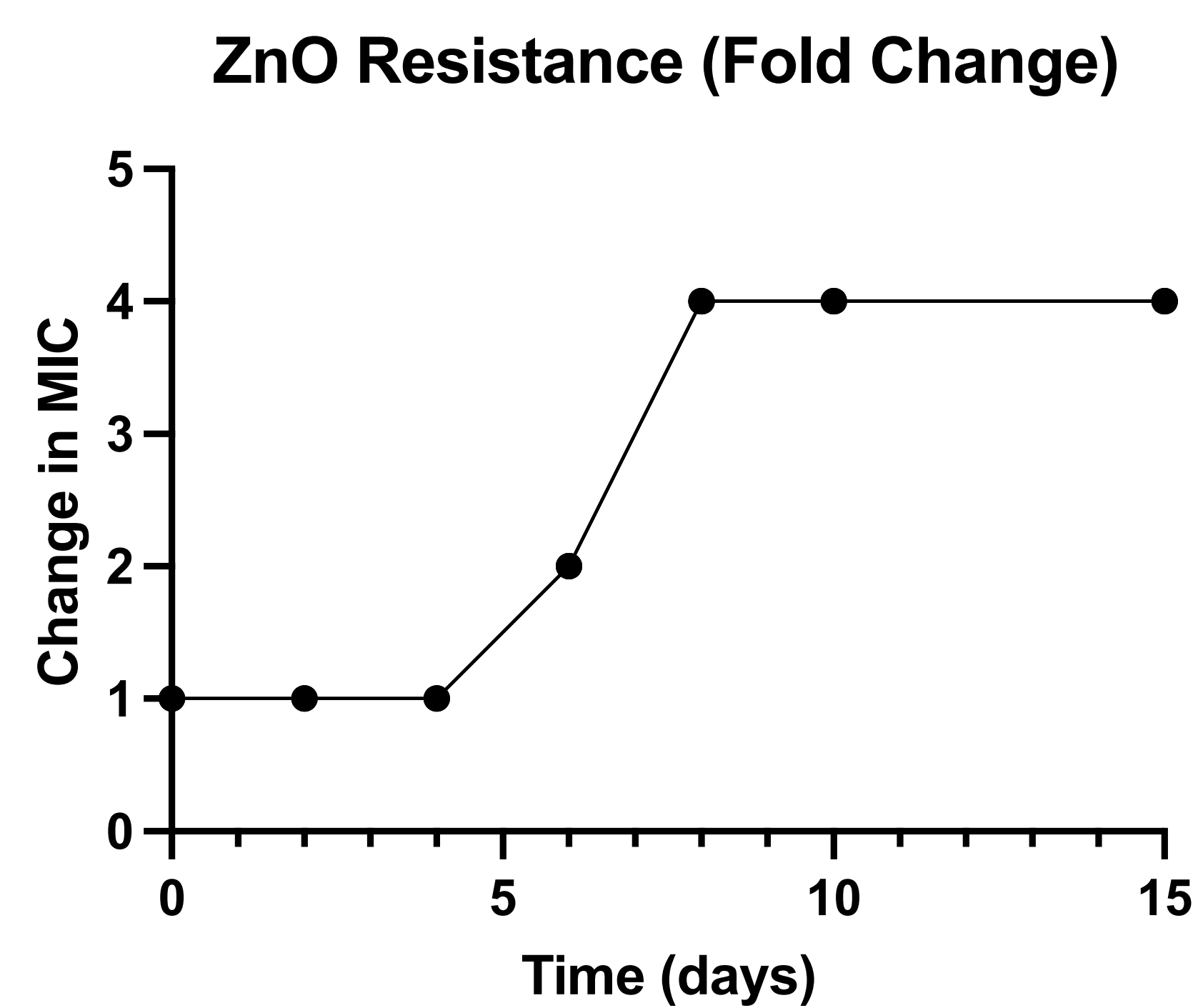
Background

Proposed Mechanisms of ZnO NP Antibacterial Activity



Literature suggests that the mechanism of ZnO antibacterial action could occur through internalization of the ZnO that results in disruptions in cellular function, production of ROS, production of Zn²⁺ ions and other toxic ionic complexes, and loss of cellular integrity due to contact or interactions between ZnO surfaces and the cell envelope. Previous work (Reeks et al. 2021) has shown that internalization of ZnO NPs is not necessary for ZnO killing, so we chose to investigate the other mechanisms.

ZnO Resistance

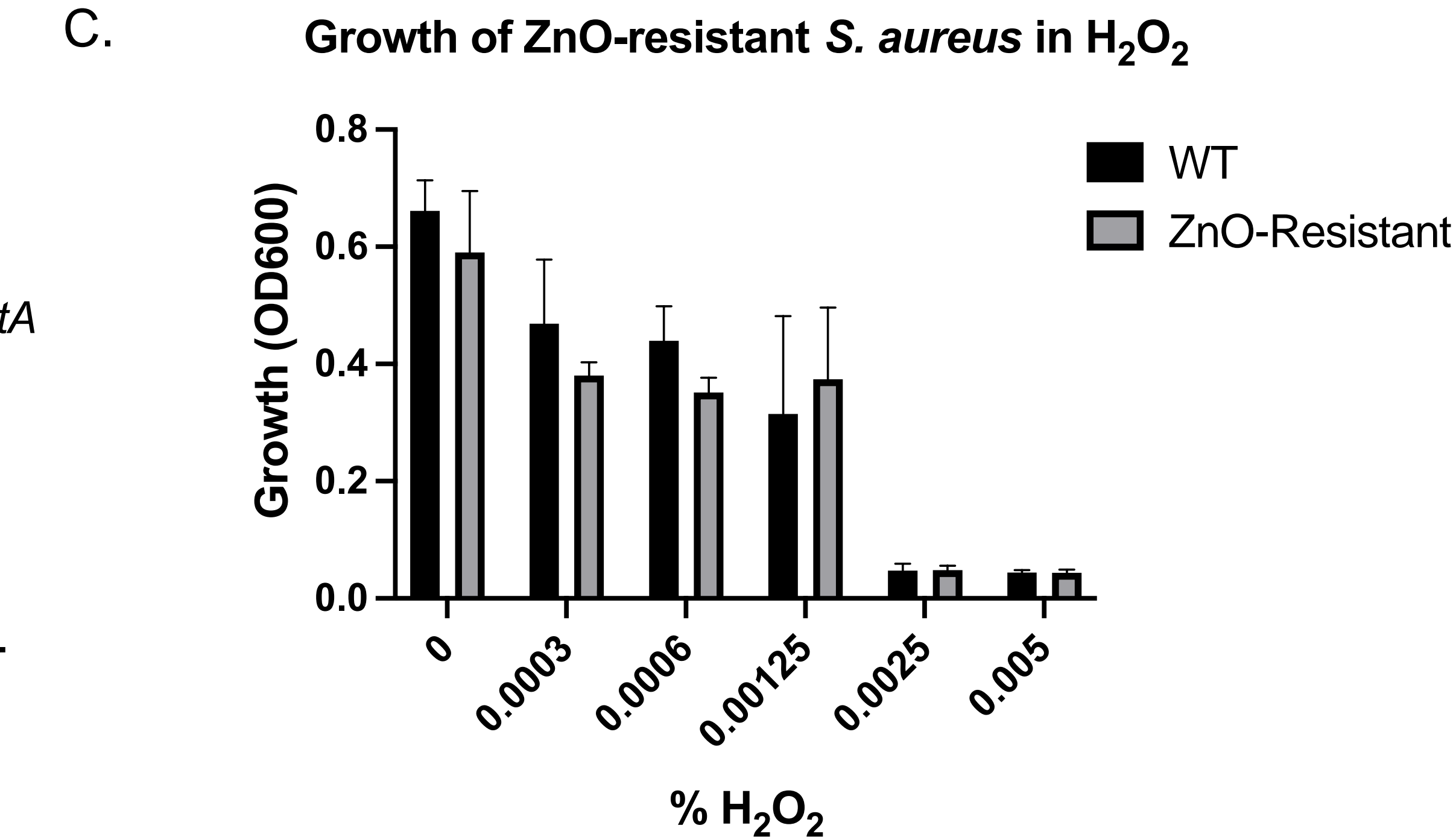
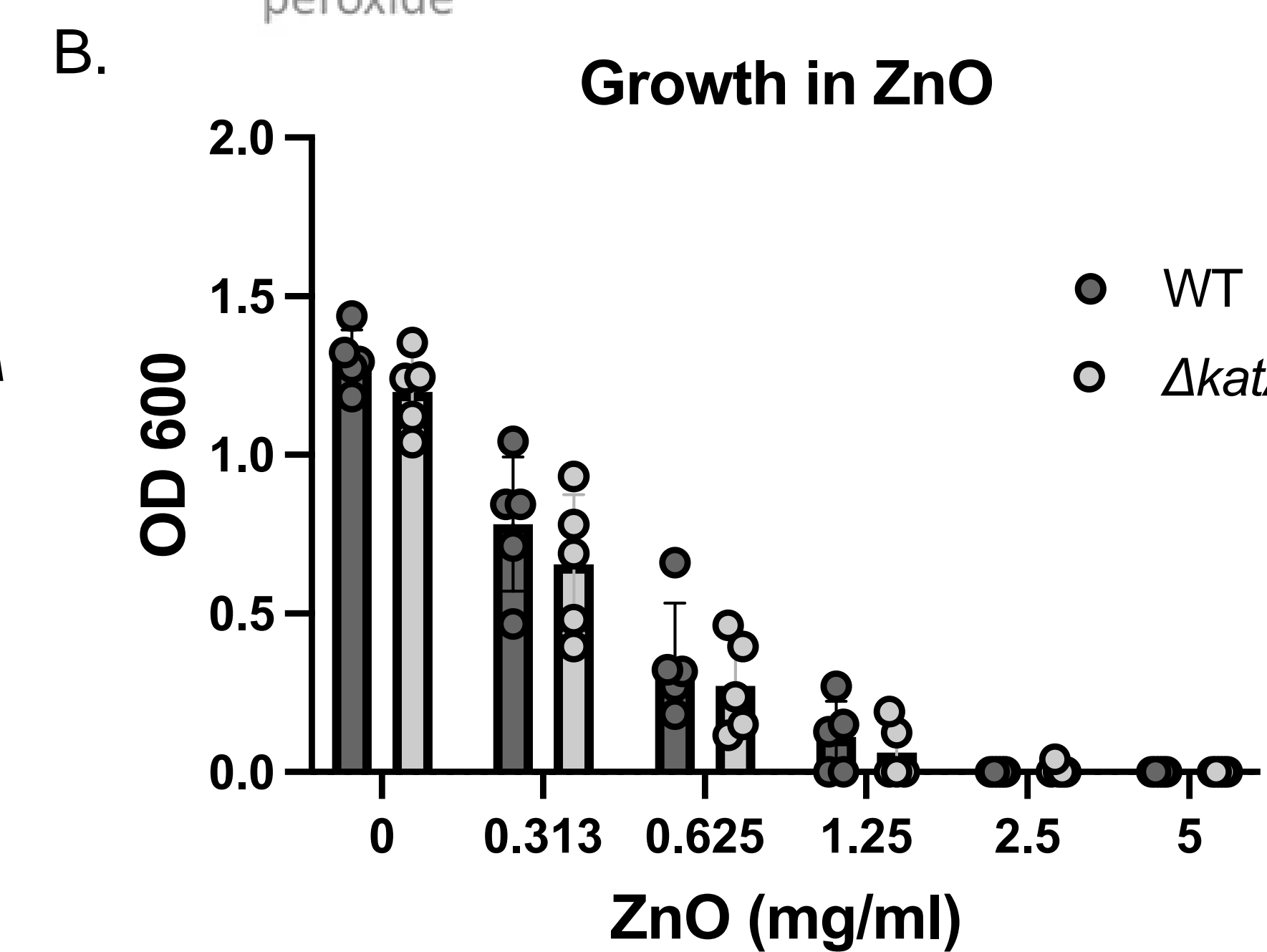
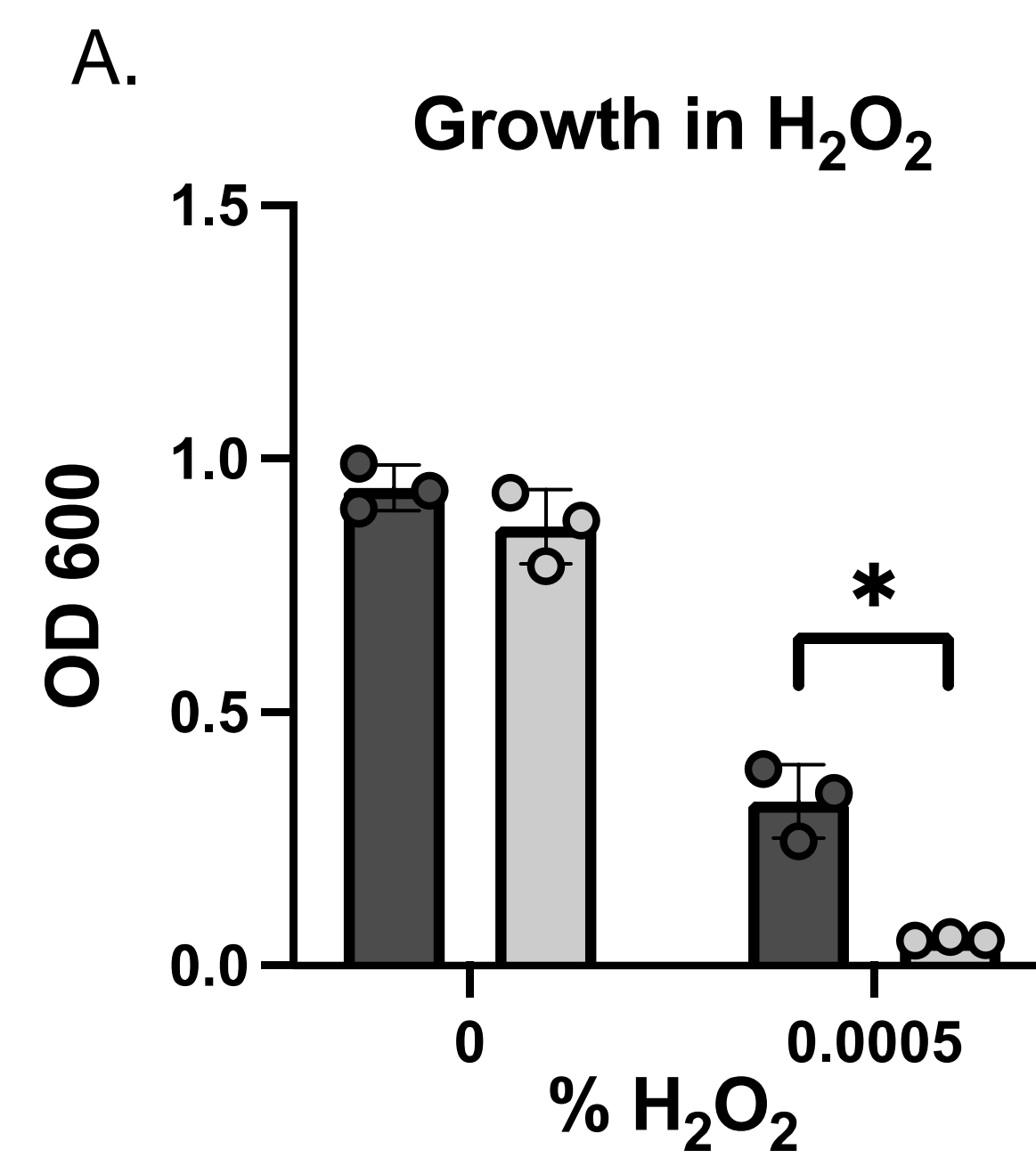


At day 0, the MIC of ZnO was 1.25 mg/ml. After 8 days, of passing the *S. aureus* at sublethal conditions, we observed a 4-fold increase in the MIC (from 1.25 mg/ml to 5 mg/ml).

Comparison of the MICs of different ZnO NP sources with wild type and ZnO Resistant *S. aureus*

Particle Type	Wt MIC (mg/ml)	ZnO ^R MIC (mg/ml)
Sigma Aldrich	1.25	5
Alfa Aesar	1.25	5
ZoChem	2.5	10

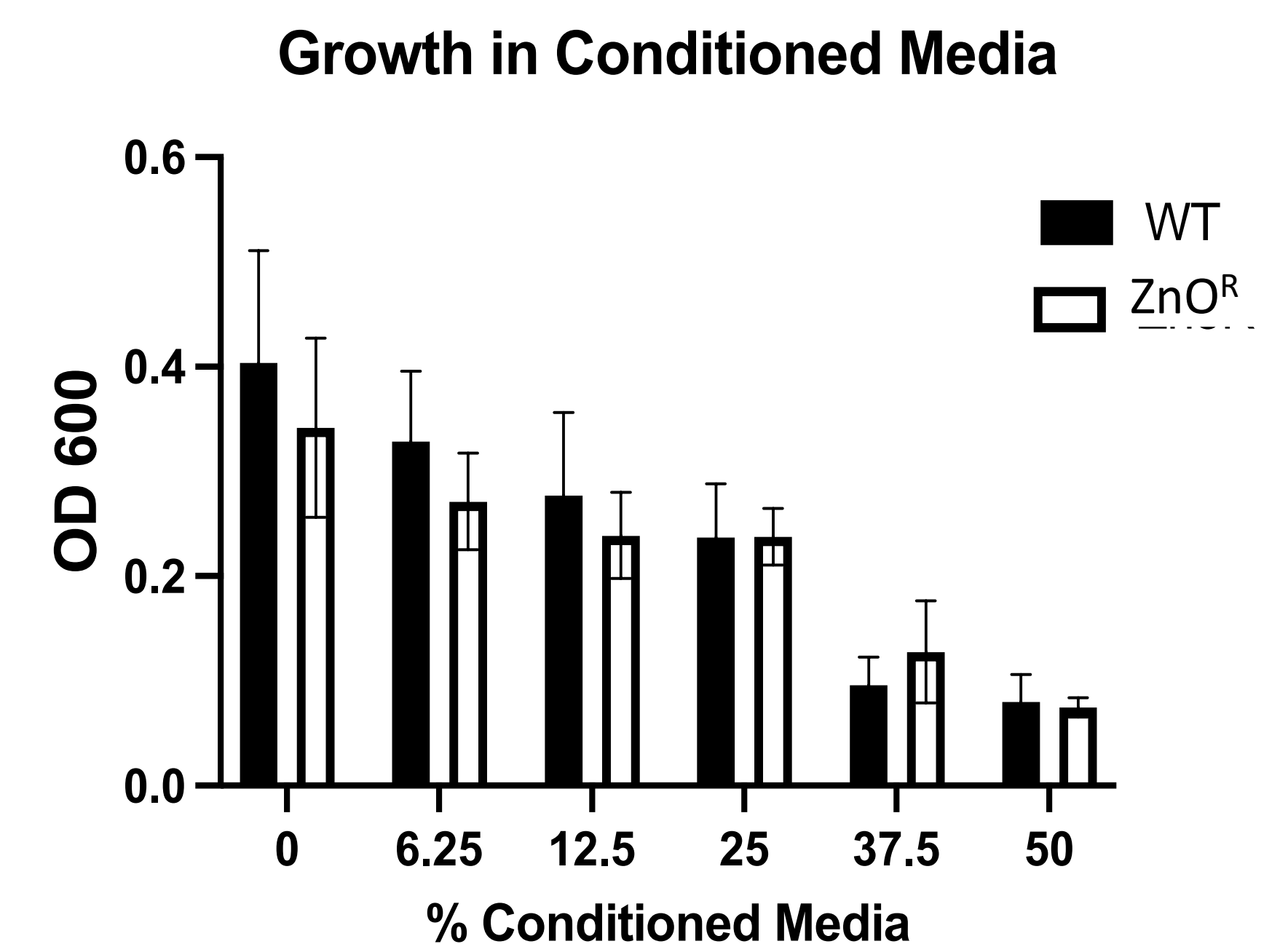
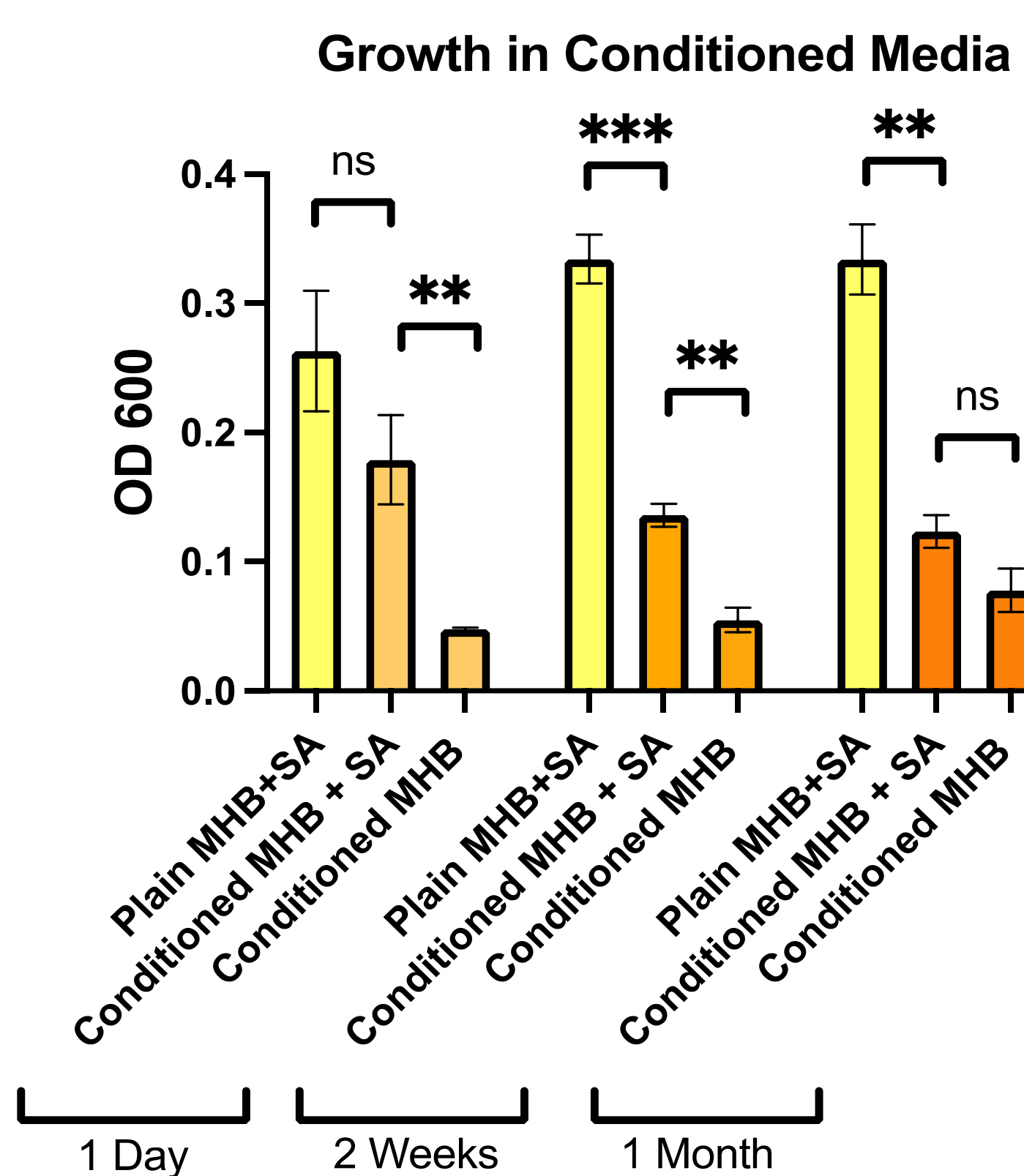
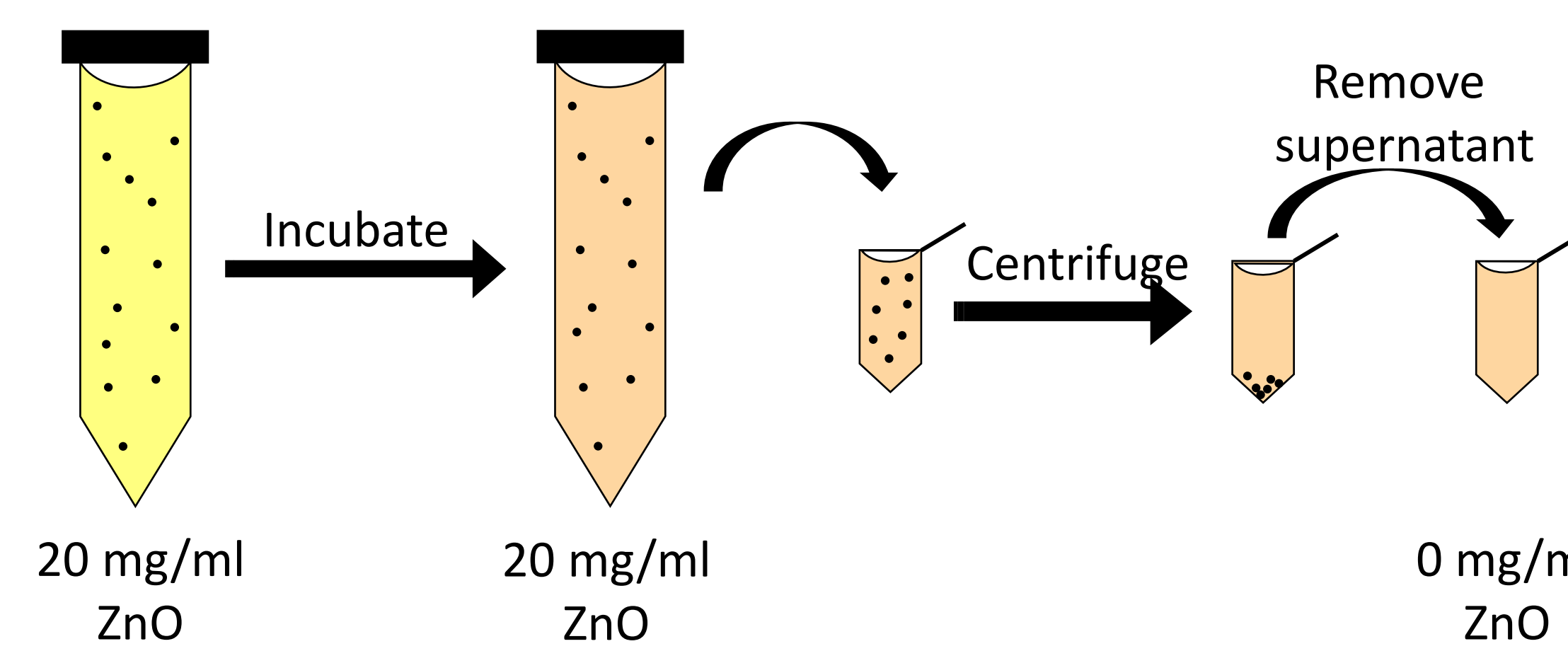
Role of H₂O₂



Production of H₂O₂ is not responsible for the antimicrobial activity of ZnO NPs. a) WT Newman and $\Delta katA$ *S. aureus* growth in H₂O₂. b) WT Newman and $\Delta katA$ *S. aureus* growth in Sigma-Aldrich NPs. c) WT Newman and ZnO Resistant *S. aureus* in H₂O₂. a-c) Error bars represent mean \pm SD of at least three independent trials. **p < 0.01 as determined by unpaired t-test.

Role of Physical Contact

Method:

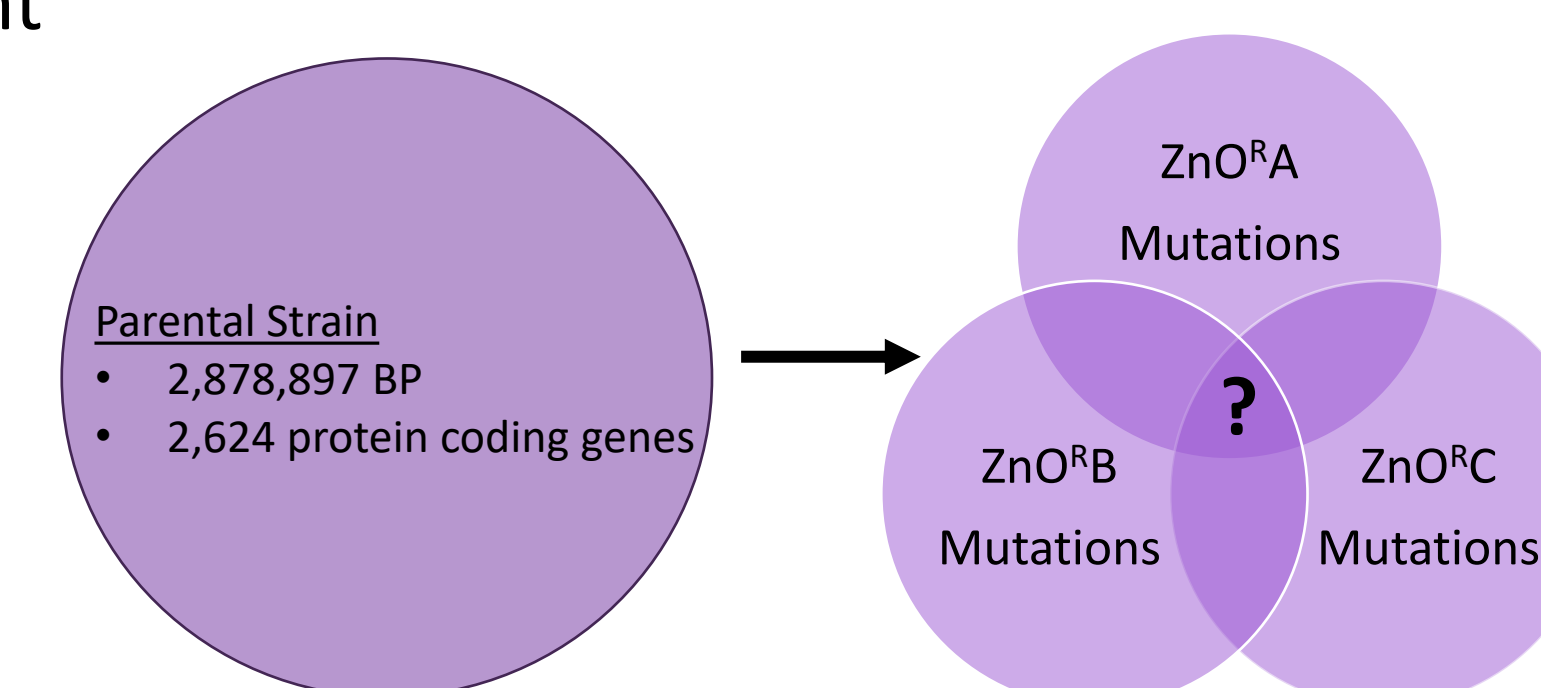


Media conditioned with ZnO NPs retains ability to inhibit bacterial growth after removal of NPs Growth of *S. aureus* was measured in normal MHB and conditioned media. Conditioned media without *S. aureus* acted as a blank. Data are presented as mean \pm SD and assays were repeated at least three independent times. Different letters represent statistically significant differences (*p < 0.05, **p < 0.01) as determined by one-way ANOVA.

Conclusions and Future Directions

1. Attempt to generate a *S. aureus* strain that is resistant to ZnO conditioned media
2. Evaluate the role of Zn²⁺ using cytochrome c assay to measure cell charge

3. WGS of Independent ZnO^R Strains



References

- John M. Reeks, Iman Ali, William J. Moss, Eric Davis, Shauna M. McGillivray, and Yuri M. Strzhemechny, "Microscale ZnO with controllable crystal morphology as a platform to study antibacterial action on *Staphylococcus aureus*", *Biointerphases* 16, 031003 (2021)
- Sirelkhatim, A., Mahmud, S., Seeni, A. et al. Review on Zinc Oxide Nanoparticles: Antibacterial Activity and Toxicity Mechanism. *Nano-Micro Lett.* 7, 219–242 (2015).
- Siddiqi, K. S., Ur Rahman, A., Tajuddin, & Husen, A. (2018). Properties of Zinc Oxide Nanoparticles and Their Activity Against Microbes. *Nanoscale research letters*, 13(1), 141.

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